

PLANT HORMONES—VIII.

COMBINED GAS CHROMATOGRAPHY-MASS SPECTROMETRY OF THE METHYL ESTERS OF GIBBERELLINS A₁ TO A₂₄ AND THEIR TRIMETHYLSILYL ETHERS

R. BINKS, J. MACMILLAN and R. J. PRYCE

Department of Organic Chemistry, The University, Bristol

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Abstract—Line diagrams of GC-MS low resolution mass spectra are presented for the methyl esters of gibberellins A₁ to A₂₄ and for the trimethylsilyl ethers of the methyl esters of the hydroxylated gibberellins A₁ to A₈, A₁₀, A₁₃, A₁₄, and A₁₆ to A₂₃. These reference spectra allow conclusive identification of the presently known gibberellins without access to authentic specimens. The utility of the mass spectra of these derivatives in structural diagnosis of the gibberellins is discussed. High resolution measurement of the composition of fragment ions in selected trimethylsilyl ethers are included.

COMBINED gas chromatography-mass spectrometry (GC-MS) has been shown to be an invaluable method of identifying known¹⁻³ and new⁴ gibberellins in plant extracts. The method is sensitive and can be applied to crude extracts¹ although more detailed information is obtained^{2,3} on partially purified fractions. A further advantage of the method is that the twenty-four known gibberellins can be conclusively identified without access to authentic specimens of the scarce gibberellins, provided reference mass spectra are available. These reference spectra are presented in Figs. 1 and 2 for the methyl esters of gibberellins A₁ to A₂₄ (MeA₁ to MeA₂₄) and for the trimethyl silyl ethers (TMSi ethers) of the methyl esters of the hydroxylated gibberellins A₁ to A₈, A₁₀, A₁₃, A₁₄, and A₁₆ to A₂₃; the utility of the mass spectra of these derivatives in structural diagnosis of the gibberellins is presented below. For convenience the numbering of the gibbane nucleus is shown in (I).

Methyl Ester Trimethylsilyl Ethers (Fig. 1)

The spectra are distinct and characteristic for all hydroxylated gibberellins including those of isomeric composition. From a comparison with corresponding methyl esters the number of hydroxyl groups can be determined. Molecular ions (M⁺) are observed in all the spectra and show the computed isotope peaks for the presence of one, two, or three silicon atoms. Other common features include fragment ions at M-31/32 and M-59/60 associated with the methoxycarbonyl group and M-15, M-89/90, *m/e* 73, and *m/e* 75, associated with the TMSi

¹ J. MACMILLAN, R. J. PRYCE, G. EGLINTON and A. MCCORMICK, *Tetrahedron Letters* 2241 (1967).

² R. J. PRYCE, J. MACMILLAN and A. MCCORMICK, *Tetrahedron Letters* 5009 (1967).

³ J. MACMILLAN and R. J. PRYCE, *Tetrahedron Letters* 1537 (1968).

⁴ R. J. PRYCE and J. MACMILLAN, *Tetrahedron Letters* 4173 (1967).

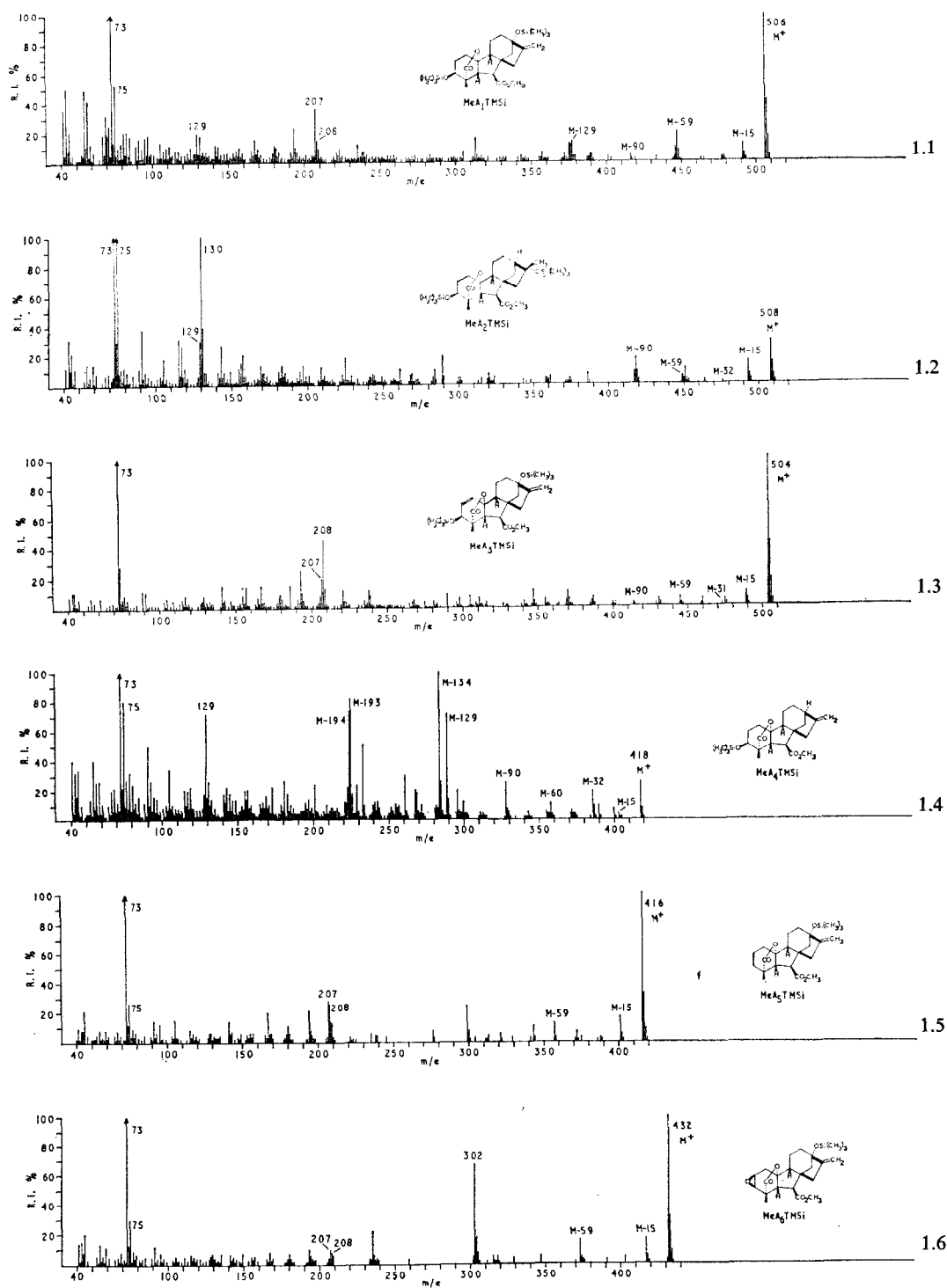


FIG. 1. MASS SPECTRA OF THE GIBBERELLIN METHYL ESTER TMSi ETHERS.

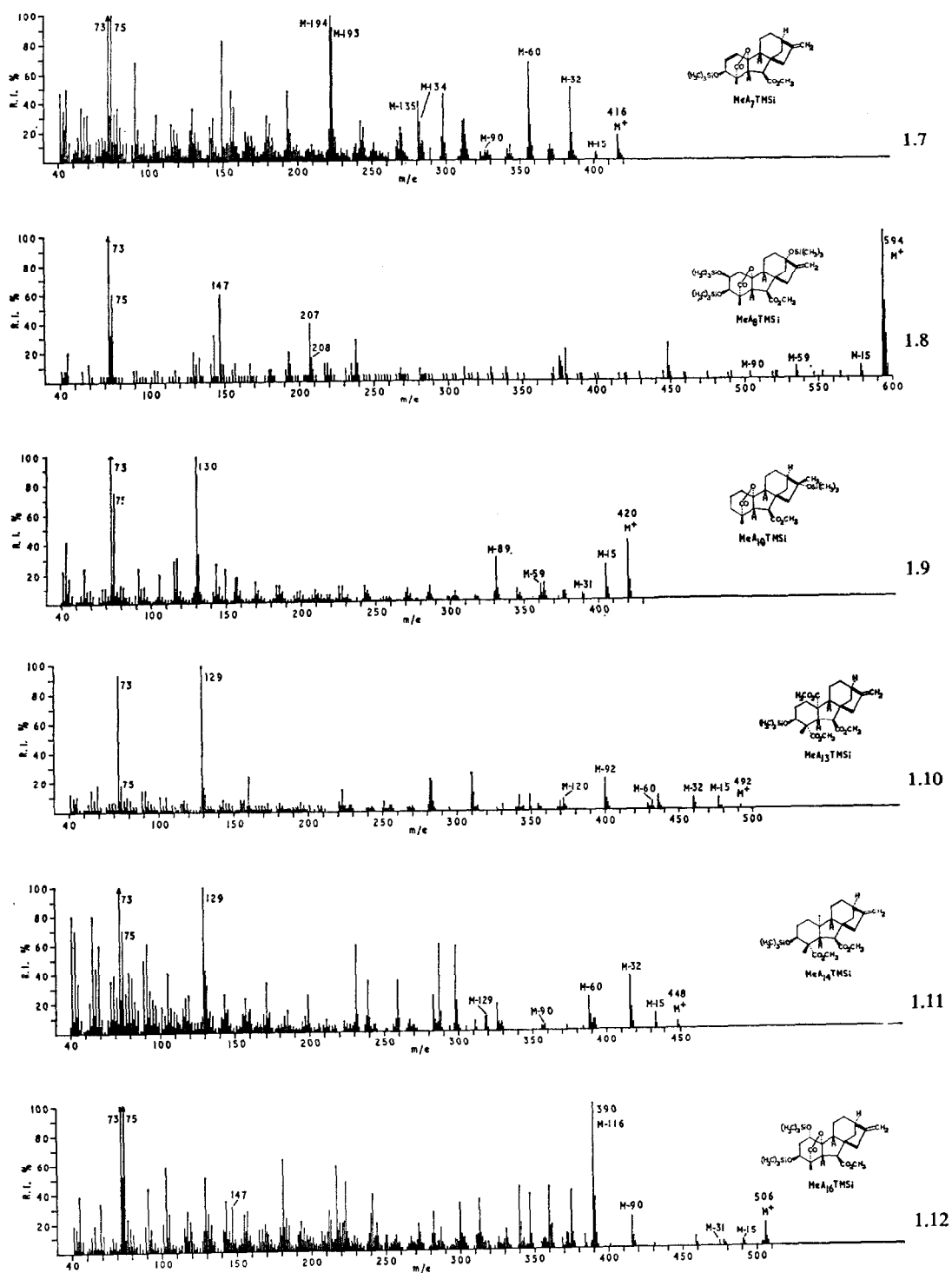


FIG. 1—continued.

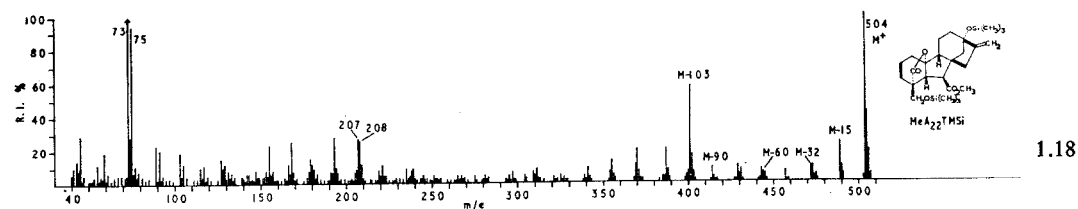
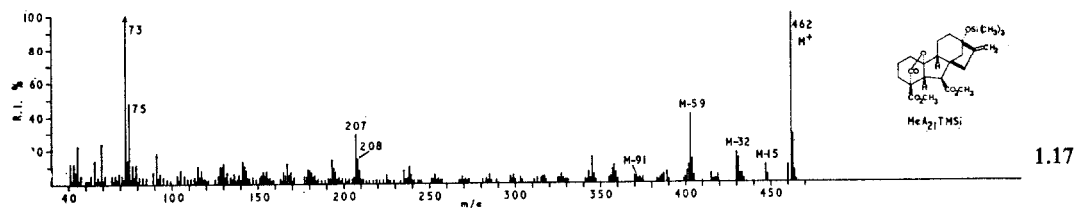
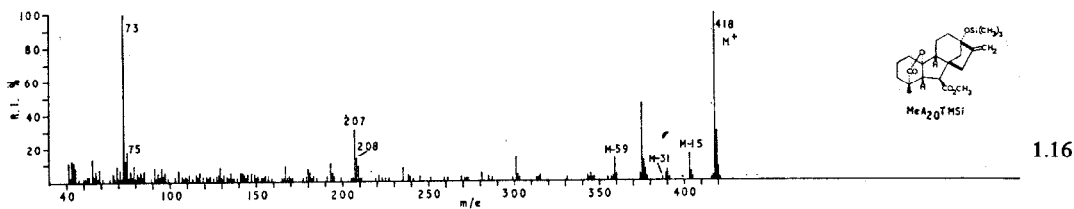
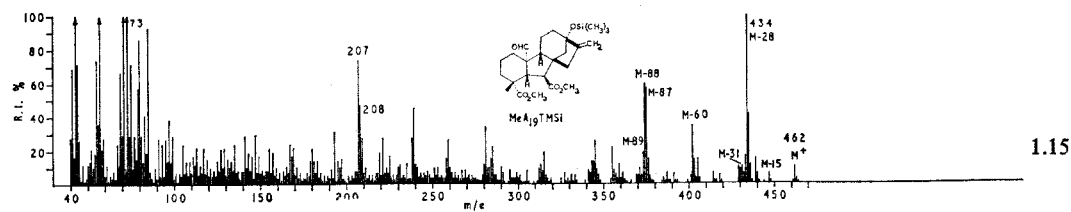
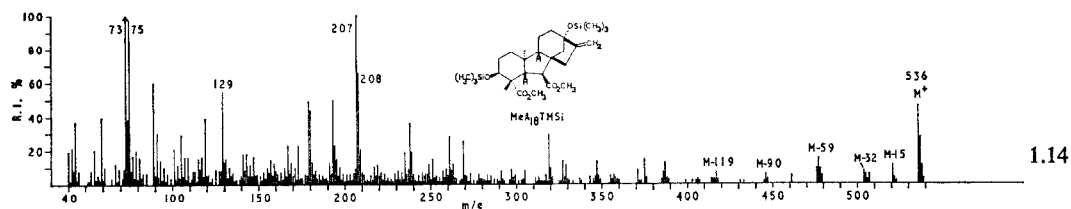
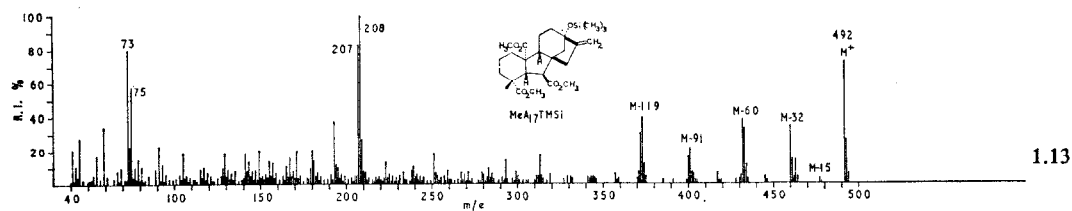


FIG. 1—continued.

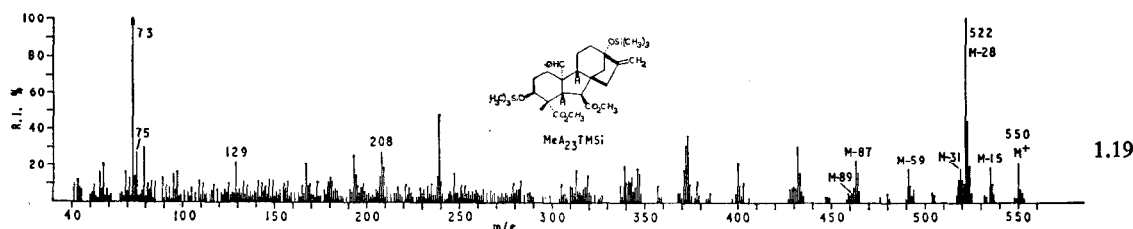
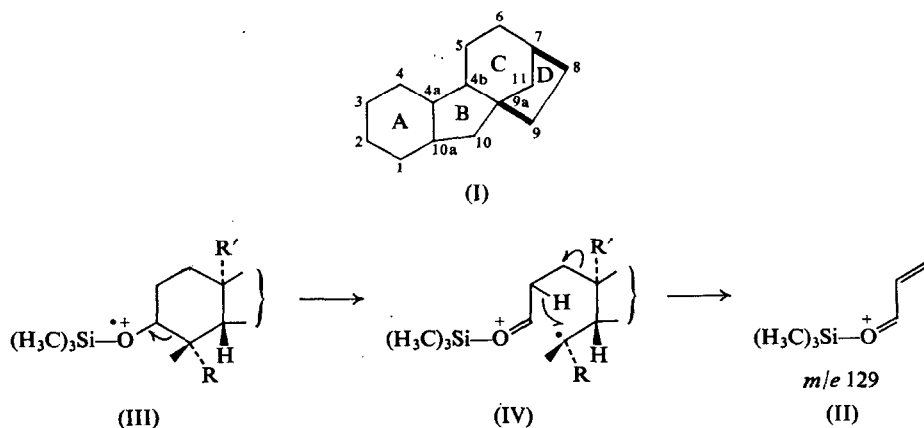


FIG. 1—continued.

ether group. In the spectra of the methyl ester TMSi ethers of the C_{20} -gibberellins and gibberellin A_{21} prominent peaks at $M-90/91/92$ are probably due to fragmentation of two methoxycarbonyl groups [$M-(50/60 + 31/32)$]. The ion at m/e 73 is very intense in all spectra and was not taken as the base peak for any of the line diagrams.

2-Hydroxygibberellins are characterized in the mass spectra of their methyl ester TMSi ethers by a prominent ion at m/e 129, shown to be $C_6H_{13}OSi$ in the case of MeA_4 TMSi. This ion, accompanied in some cases by an $M-129$ peak, presumably has the structure (II) arising from ring A by the process (III) \rightarrow (IV) \rightarrow (II) which is typical of such TMSi ethers.⁵

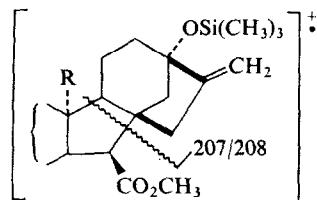


The TMSi ethers of 7-hydroxygibberellin methyl esters are distinguished by an intense molecular ion which is the base peak in all the 7-TMSi ethers except those of gibberellins A_{17} , A_{18} , A_{19} , and A_{23} . In contrast, the molecular ions of the TMSi ethers of the 7-deoxygibberellins are of much lower intensity. This correlation is clearly illustrated by comparison of the isomeric TMSi ethers of gibberellins A_{13} (Fig. 1.10) and A_{17} (Fig. 1.13) methyl esters. This comparison also suggests a further distinction between these isomeric 2- and 7-mono-hydroxy- C_{20} -gibberellins, namely that the $M-119/120$ peak (see discussion of methyl ester spectra below) is stronger in the absence of the 2-TMSi ether grouping. Significant peaks at m/e 207/208, shown to be $C_{12}H_{19/20}OSi$ in the case of MeA_5 TMSi, are characteristic of the 7-TMSi ethers; they must contain rings C and D and probably arise by the cleavage shown in (V) with or without additional loss of hydrogen.

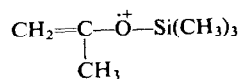
The methyl ester TMSi ethers of gibberellins A_2 (Fig. 1.2) and A_{10} (Fig. 1.9) have intense base peaks at m/e 130. This ion, shown to have the composition $C_6H_{14}OSi$ in the case of

⁵ J. DIEKMAN and C. DJERASSI, *J. Org. Chem.* **32**, 1005 (1967).

MeA₁₀ TMSi, is probably (VI) from cleavage of ring D and, therefore, diagnostic of an 8-TMSi ether.

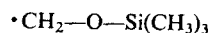


(V)

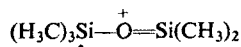
 m/e 130

(VI)

The TMSi ether of the 1-hydroxymethyl group in gibberellin A₂₂ is characterized by a peak at M-103 (Fig. 1.18); this ion corresponds to the loss of the fragment (VII).

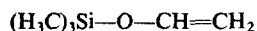


(VII)

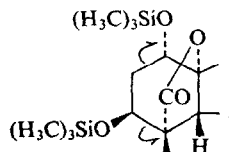
 m/e 147

(VIII)

The vicinal 2,3-bis-TMSi ether of MeA₈ (Fig. 1.8) gives rise to an ion m/e 147 corresponding to (VIII); since this ion is observed in a spectrum obtained by GC-MS, it must come from rearrangement of the vicinal TMSi ether and cannot be the artefact from hexamethyldisilazane or trimethylsilyl chloride described by Budzikiewicz *et al.*⁶ The bis-TMSi ether of MeA₁₆ (Fig. 1.12) is unique in showing a base peak at M-116 (C₂₁H₃₀O₅Si) which could arise by loss of the fragment (IX) from ring A by the process suggested in (X).



(IX)



(X)

The TMSi ethers of MeA₁₉ and MeA₂₃ (Figs. 1.15 and 1.19 respectively) show an intense (base) peak at M-28 corresponding to the loss of CO from the aldehyde function. These two TMSi ethers also show prominent peaks at M-87/88 which are probably due to M-(28 + 59/60). Peaks of low intensity at M-28/29 are also observed in the spectra of the TMSi ethers of several non-aldehydic gibberellins but their origin is unknown.

The spectra of MeA₄ TMSi (Fig. 1.4) and MeA₇ TMSi (Fig. 1.7) are distinguished by intense peaks at M-134/135 (M-134 was shown to be C₁₅H₂₈O₃Si in the case of MeA₄ TMSi and C₁₅H₂₆O₃Si in the case of MeA₇ TMSi; M-135 was shown to be C₁₅H₂₅O₃Si in the case of MeA₇ TMSi), and at M-193/194 (C₁₃H_{24/25}O₅Si in the case of MeA₄ TMSi and C₂₃H_{22/23}O₅Si in the case of MeA₇ TMSi). These fragment ions must contain the A rings of MeA₄ and MeA₇ TMSi ethers. Further, the M-134/135 and M-193/194 ions differ in composition by C₂H₃O₂ which could correspond to loss of a methoxycarbonyl group and suggests that the M-134/135 ions also contain the C-10 and the 10-methoxycarbonyl group.

⁶ H. BUDZIKIEWICZ, C. DJERASSI and D. H. WILLIAMS, *Mass Spectrometry of Organic Compounds*, p. 476, Holden-Day, New York (1967).

High resolution measurements on the methyl ester TMSi ethers were carried out on an MS9 instrument by direct insertion of a portion of each reaction mixture of gibberellin methyl ester TMSi ether together with the silylating reagent (hexamethyldisilazane, trimethylsilyl chloride, and pyridine). During this work intense peaks arising from the silylating reagent were observed at m/e 130 ($C_4H_{12}NSi_2$) and m/e 147 ($C_5H_{15}OSi_2$). Only the latter of these two peaks has previously been noted⁶ as being derived from the silylating reagent.

Methyl Esters (Fig. 2)

The mass spectra of alkyl esters of several C_{19} -gibberellins have been reported by two groups. Wulfson *et al.*^{7,8} have discussed the low resolution spectra of the methyl and ethyl esters of gibberellins A_1 , A_3 , A_4 , and A_7 while the high resolution spectra of the methyl

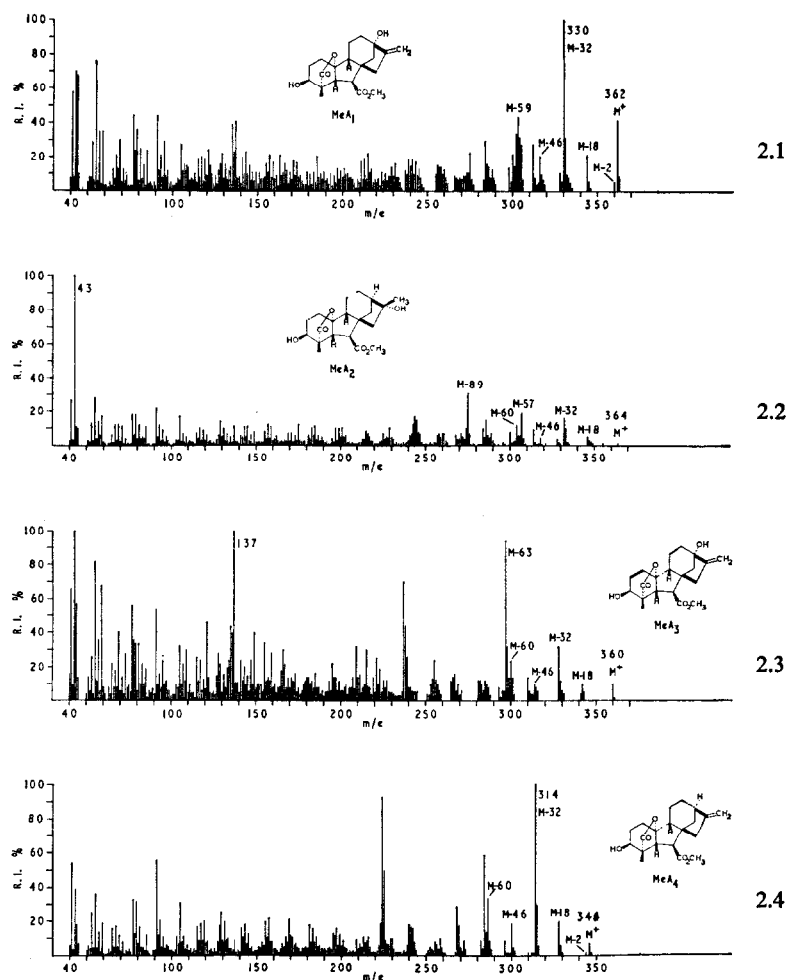


FIG. 2. MASS SPECTRA OF THE GIBBERELLIN METHYL ESTERS.

⁷ N. S. WULFSON, V. I. ZARETSKII, I. B. PAPERNAJA, E. P. SEREBRYAKOV and V. F. KUCKEROV, *Tetrahedron Letters* 4209 (1965).

⁸ V. I. ZORETSKII, N. S. WULFSON, I. B. PAPERNAJA, I. A. GURVICH, V. F. KUCHEROV, I. M. MILSTEIN, E. P. SEREBRYAKOV and A. V. SIMOLIN, *Tetrahedron* 24, 2327 (1968).

esters of gibberellin A₁ to A₅, A₇ to A₉, and A₂₀ have been discussed by Takahashi *et al.*⁹ Some high resolution spectral data has been presented for the methyl esters of gibberellins A₁₉,¹⁰ A₂₁,¹¹ A₂₂,¹¹ and A₂₄¹² and there are some low resolution data available for gibberellin A₂₃¹³ methyl ester. In general the present data confirm and extend the conclusions of the above workers although some modifications to their correlation of fragmentation patterns with structure are suggested.

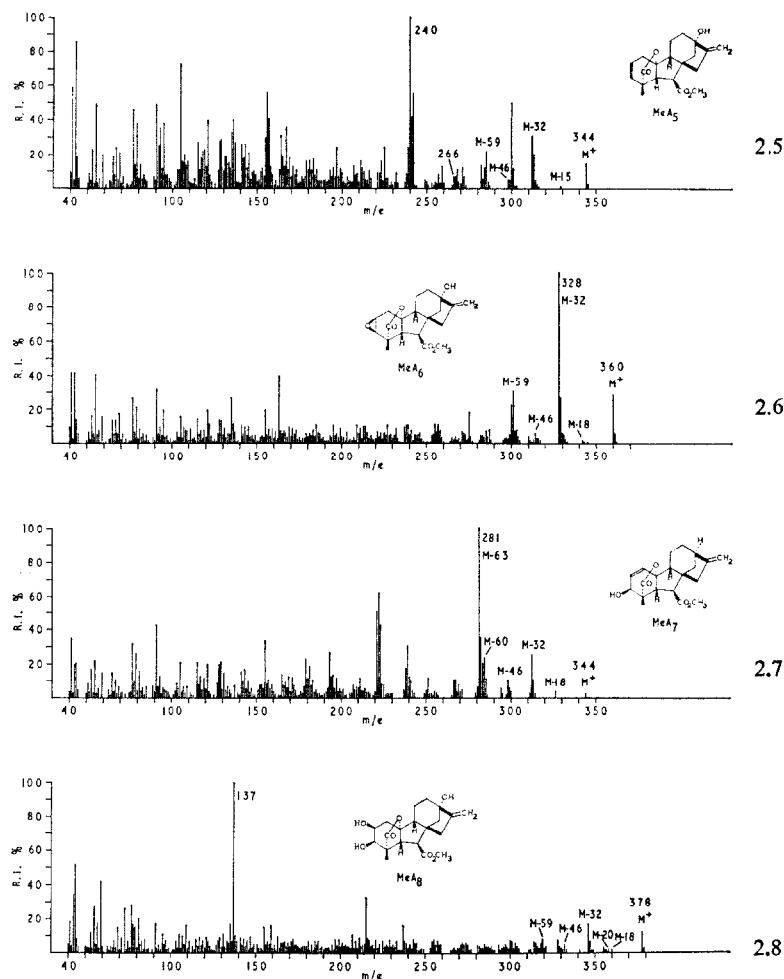


FIG. 2—continued.

⁹ N. TAKAHASHI, N. MUROFUSHI, S. TAMURA, N. WASADA, H. HOSHINO, T. TSUCHIYA, T. AOYAMA and H. MORITA, *Tetrahedron Letters* 895 (1967).

¹⁰ N. MUROFUSHI, S. IRIUCHIJIMA, N. TAKAHASHI, S. TAMURA, J. KATO, Y. WADA, E. WATANABE and T. AOYAMA, *Agr. Biol. Chem. (Tokyo)* 30, 917 (1966).

¹¹ N. TAKAHASHI, N. MUROFUSHI, T. YOKOTA, S. TAMURA, J. KATO and Y. SHIOTANI, *Tetrahedron Letters* 4861 (1967).

¹² D. M. HARRISON, J. MACMILLAN and R. H. B. GALT, *Tetrahedron Letters* 3137 (1968).

¹³ K. KOSHIMIZU, H. FUKUI, M. INUI, Y. OGAWA and T. MITSUI, *Tetrahedron Letters* 1143 (1968).

The fragmentation of the methyl esters is less clearly diagnostic of structure than in the TMSi ethers. The loss of 31 and 32 and 59 and 60 mass units from the molecular ion, observed by the previous workers, is characteristic of methoxycarbonyl groups and is not very informative. The methyl esters of C_{20} -gibberellins possess two or three methoxycarbonyl groups and

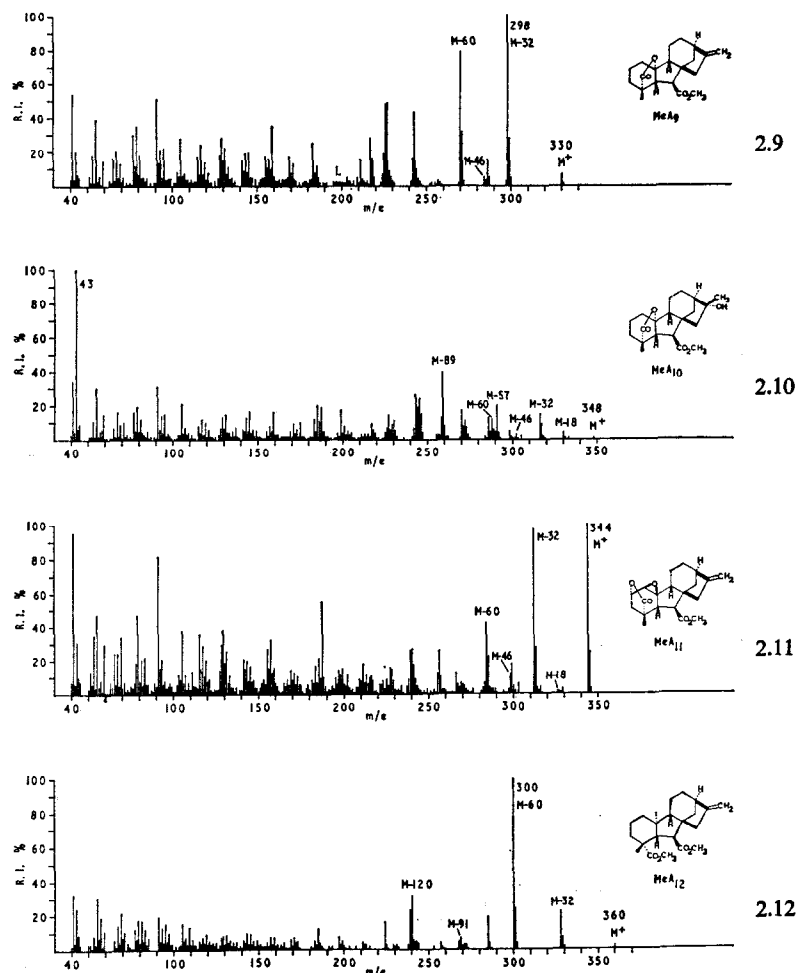


FIG. 2—continued.

can be distinguished from the C_{19} -gibberellins, with the exception of the dicarboxylic acid gibberellin A₂₁, by intense peaks at M-119/120 in addition to the M-59/60 peak. A 2-hydroxyl group appears to inhibit the loss of the second 60 mass units since MeA₁₇ (Fig. 2.17) shows a more intense M-120 ion than MeA₁₃ (Fig. 2.13). Similar relative intensities in the mass spectra of MeA₁₇ TMSi and MeA₁₃ TMSi (see discussion of MeTMSi spectra above and Figs. 1.13 and 1.10 respectively). While the methyl esters of all the C_{20} -gibberellins with two or three methoxycarbonyl groups all show distinct M-119/120 peaks only gibberellins A₁₃, A₁₇, and A₁₈ have such peaks in the mass spectra of the MeTMSi/s. The mass spectra of the methyl

esters of these di- and tri-carboxylic acids (the C₂₀-gibberellins and gibberellin A₂₁) also show peaks at M-91/92 which presumably arise from loss of 59/60 and 31/32 mass units from two methoxycarbonyl groups. MeA₂₃ (Fig. 2.23) shows an intense peak at M-64 presumably due to M-(2 × 32) from two methoxycarbonyl groups. The dimethyl esters of gibberellins

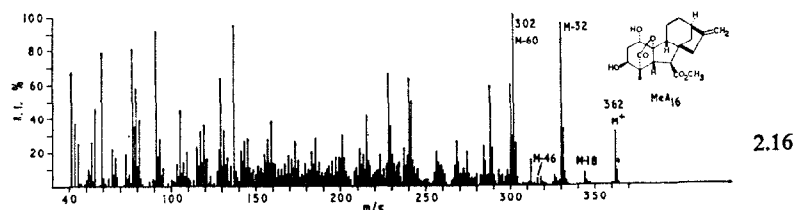
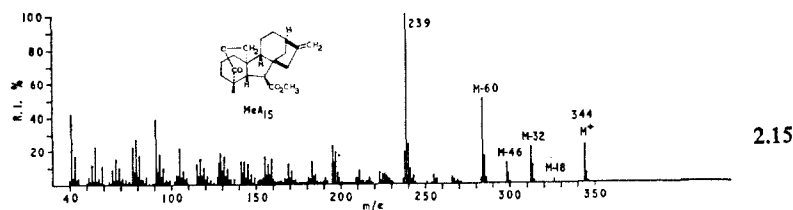
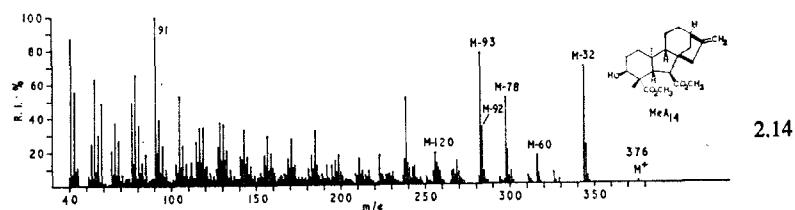
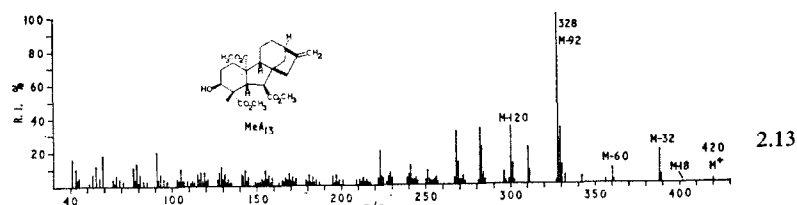


FIG. 2—continued.

A₁₄ (Fig. 2.14) and A₁₈ (Fig. 2.18) show very similar fragmentation patterns; the characteristic peaks at M-60, M-78, and M-93 possibly representing the successive losses of methyl formate, water, and a methyl group; in these two esters the M-93 peak is much more intense than the M-92 peak.

An M-18 peak is shown not only by the hydroxylated gibberellins but also by the two epoxides, MeA₆ (Fig. 2.6) and MeA₁₁ (Fig. 2.11) and, unexpectedly by the MeA₁₅ (Fig. 2.15). The relative intensities of the M-18 peaks in the isomeric gibberellins MeA₁₃ and MeA₁₇

have been measured accurately both in GC-MS spectra obtained with an LKB 9000 instrument and in spectra obtained by direct insertion of the solids using an A.E.I. MS9 spectrometer, at an ionization voltage of 70 eV in both cases. MeA₁₇ showed an M-18 peak with 30 and 80 per cent of the molecular ion in the respective instruments, compared with 20 and

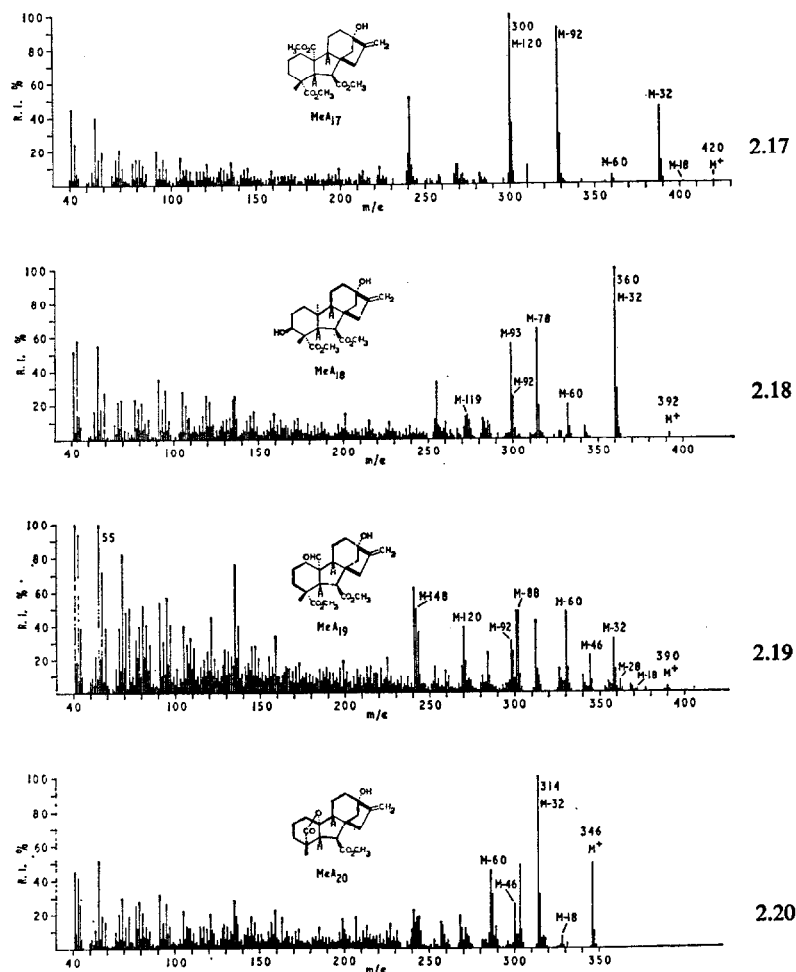


FIG. 2—continued.

17 per cent for MeA₁₃. These results contradict a previous suggestion⁹ that 7-hydroxy-gibberellins can be distinguished from gibberellins with a 2-hydroxyl group by the lower intensity of the M-18 peak. The location of the hydroxyl group in gibberellins is most securely established from the spectra of the MeTMSi derivatives.

The methyl esters of gibberellins A₁ (Fig. 2.1) and A₄ (Fig. 2.4) show an M-2 peak which must be due to dehydrogenation of the 2-hydroxyl function and not to incomplete separation of MeA₃ from MeA₁ by GC or to the presence of MeA₄ and MeA₅ in gibberellin A₄ methyl ester. The spectrum of MeA₁ does not contain peaks at m/e 342 and 297 characteristic of

MeA₃ (Fig. 2.3); similarly the spectrum of MeA₄ does not contain peaks at m/e 281 (MeA₇, Fig. 2.7) or 266 (MeA₅, Fig. 2.5). The peak shown by MeA₈ (Fig. 2.8) at M-20 is possibly the loss of 18 mass units from an M-2 ion.

The M-46 peak, previously assigned⁹ to the loss of CO₂ and H₂ from the C₁₉-gibberellins with a five-ring lactone, is also shown by MeA₁₅ (Fig. 2.15) with a six-ring lactone. However, an M-46 peak is also shown by the methyl esters of the aldehydic gibberellins A₁₉

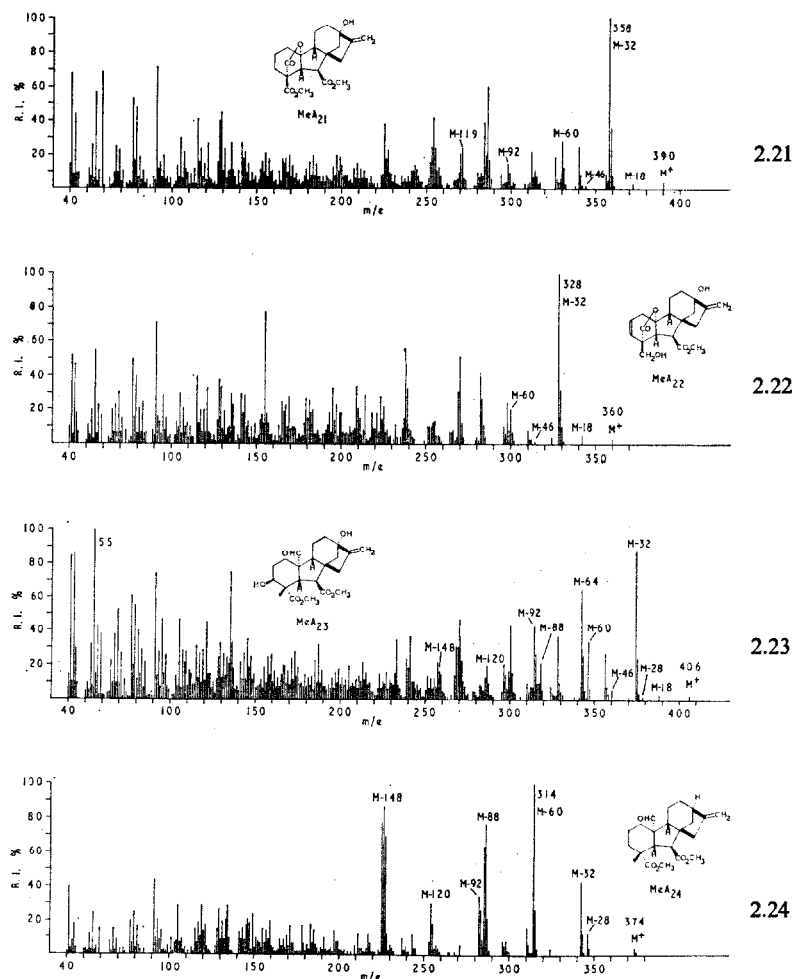
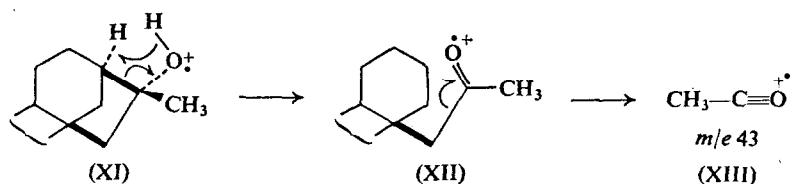


FIG. 2—continued.

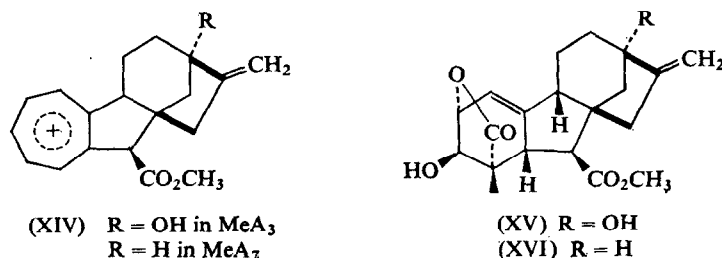
(Fig. 2.19), A₂₃ (Fig. 2.23), and A₂₄ (only 0.5 per cent of base peak and not shown in Fig. 2.24) and possibly represents the combined loss of 18 and 28 mass units. The methyl esters of these aldehydic gibberellins are characterized by the loss of 28 mass units from the molecular ion and from the M-32 (M-60), M-60 (M-88) and M-120 (M-148) ions.

The methyl esters of gibberellins A₂ and A₁₀ (Figs. 2.2 and 2.10 respectively) show identical fragmentation patterns with a base peak at m/e 43 which could arise by the fragmentation pathway (XI) → (XII) → (XIII). The peak at M-57 is considered to represent the cleavage



of ring D with the transfer of one hydrogen atom and the peak at M-89 to represent the same cleavage combined with loss of 32 mass units.

The mass spectra of MeA₃ (Fig. 2.3) and MeA₇ (Fig. 2.7) are characterized by a very intense peak at M-63. This peak probably represents the loss of H₂O, CO₂, and H from ring A presumably to give a tropylium ion (XIV). This result is contrary to the findings of Wulfson *et al.*⁸ who reported that MeA₃ and MeA₇ showed intense peaks at M-62 and could be



distinguished from their corresponding isomeric 1→3-lactones (XV) and (XVI) which showed intense M-63 peaks. It is our experience that the mass spectra of MeA₃ and its isomeric 1→3-lactone (XV), and of MeA₇ and its isomeric 1→3-lactone (XVI), are virtually indistinguishable.

Whilst we have often observed M + 14 peaks in mass spectra obtained by direct insertion of gibberellin methyl esters prepared with diazomethane, no such peaks were observed in the GC-MS mass spectra. Diazomethane is known to form a cyclopropyl derivative with the exocyclic double bond in gibberellin A₄.¹⁴

EXPERIMENTAL

Low resolution mass spectra were obtained with an LKB 9000 combined gas chromatograph-mass spectrometer operating at 70 eV. GC columns used were 6 ft × $\frac{1}{8}$ in. i.d. (glass) packed with 1 or 2 per cent QF-1 on acid-washed and silanized Gas Chrom P or A respectively, and 10 ft × $\frac{1}{8}$ in. i.d. (glass) packed with 1 per cent SE-30 on acid-washed and silanized Gas Chrom P. For both types of GC columns operating temperatures were ca. 200° with a helium flow rate of ca. 30 ml/min. The flash heater and molecular separator were both kept at ca. 250° for all determinations. The scan time for each mass spectrum was between 4 and 6 sec. For each mass spectrum the background mass spectra of SE-30 and QF-1 were subtracted as appropriate.

Samples of the gibberellin methyl esters and methyl ester TMSi ethers were obtained as previously described.¹⁵ Each mass spectrum was obtained from GC of ca. 5 μg of sample which represents a convenient, but not minimal, quantity facilitating subtraction of background spectra. GC-MS of the methyl esters was carried out exclusively on the QF-1 columns where decomposition was minimal,¹⁵ and GC-MS of the methyl ester TMSi ethers was carried out using either QF-1 or SE-30 columns as convenient.

High resolution mass spectral data on the gibberellin methyl ester TMSi ethers were obtained with an A.E.I. MS9 instrument by direct insertion of the evaporated silylation mixture into the ion source (70 eV and

¹⁴ D. C. ALDRIDGE, J. R. HANSON and T. P. C. MULHOLLAND, *J. Chem. Soc.* 3539 (1965).

¹⁵ B. D. CAVELL, J. MACMILLAN, R. J. PRYCE and A. C. SHEPPARD, *Phytochem.* 6, 867 (1967).

110°). Two intense peaks derived from the silylating reagent (hexamethyldisilazane and trimethylsilyl chloride in pyridine) were observed at m/e 130·049 ($C_4H_{12}NSi_2$ requires 130·050) and m/e 147·065 ($C_5H_{15}OSi_2$ requires 147·066).

Acknowledgements—We thank the S.R.C. for a Research Studentship to R.J.P. and the A.R.C. for a Research Grant. Most of the GC-MS data were recorded using the LKB 9000 instrument purchased from S.R.C. Grant B/SR/2398 to Drs. C. J. W. Brooks and G. Eglinton (Chemistry Department, University of Glasgow) with the expert help of Misses H. Humphrys and J. Malcolm. Other GC-MS data were recorded on LKB 9000 instruments at LKB Instruments Ltd., Stockholm and Croydon, and at the Meat Research Institute, Langford, near Bristol. To all these people who allowed us time on their instruments we are most grateful. We are particularly grateful to Dr. A. McCormick for his invaluable advice and guidance. Gibberellin samples were generously donated by Professors N. Takahashi and K. Koshimizu, and Drs. B. E. Cross, R. H. B. Galt, and J. R. Hanson.